

Characterization of 10 tobacco vein banding mosaic virus isolates from China

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Summary. – The complete RNA genome sequences of 10 tobacco vein banding mosaic virus (TVBMV) isolates from China were determined using five overlapping cDNA clones. TVBMV was divided into three groups for HC-Pro and CP and into two groups for P3 and 6K1. With more isolate sequences analyzed, the phylogenetic results suggest that TVBMV could be divided into four subgroups based on HC-Pro and CP and into three subgroups with P3 and 6K1. Nucleotide sequence diversity analysis showed geographical differentiation among the TVBMV isolates. Three of the 10 isolates were found to have undergone recombination and new recombination sites were identified in the TVBMV genome. All coding genes were under negative selection and the population appeared to have remained stable over a long period. This study also provides preliminary data on the 3'-untranslated region as potentially the best genome sequence for developing transgenic tobacco.

Keywords: tobacco vein banding mosaic virus; complete sequence; phylogenetic analysis; recombination; selection pressure

Introduction

TVBMV, which was discovered in Taiwan in 1964 (Chin, 1966; Tian *et al.*, 2007), is a distinct member of the genus *Potyvirus* (Chang *et al.*, 1994; Habera *et al.*, 1994). The genome of TVBMV is a single-stranded positive-sense RNA with 9570 nucleotides (nt). Its large open reading frame (ORF) is predicted to encode a polyprotein of 3079 amino acids (M_p , 348.6 kDa), which is cleaved into 10 mature proteins.

It mainly infects solanaceous plants and is transmitted by aphids in a nonpersistent manner, resulting in vein clearing, vein banding and necrotic lesions on leaves (Chin, 1975; Habera *et al.*, 1994; Zhang and Li, 2001). Its filamentous particles measure approximately 780 nm×15 nm.

TVBMV was once a major threat to tobacco production activities in North America and Taiwan, but it has rarely occurred in most regions of China (Reddick *et al.*, 1992;

Tian *et al.*, 2007; Yu *et al.*, 2007). Due to the lack of TVBMV-resistant tobacco (Zhang *et al.*, 2011), TVBMV may potentially spread quickly and cause serious damages to tobacco production in commercial planting areas. This conclusion is further supported by findings that TVBMV is becoming significantly more severe, thereby requiring greater attention (Tian *et al.*, 2007).

As evolutionary genetic analysis plays an important role in the efficient control of viral diseases, the variability of RNA viruses has been thoroughly investigated in recent years (Moreno *et al.*, 2004; Tsompana *et al.*, 2005; Tomitaka and Ohshima, 2006). However, research on TVBMV is very limited, with the scarce data thus preventing analysis of evolution of the entire coding regions of TVBMV. Specifically, the HC-Pro, P3, 6K1, and CP genes have been only moderately studied (Tian *et al.*, 2007; Zhang *et al.*, 2011). Complete genomes of only two isolates have been reported (Wang *et al.*, 2010; Yu *et al.*, 2007). The TVBMV isolates could be divided into three and two groups based on the analysis of HC-Pro and CP and on that of P3 and 6K1, respectively. Analysis of the CP genes of subgroup I strains suggests that they can be further divided into two groups, but the between-group and within-group genetic distances do not completely support such subgrouping.

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Abbreviations: TVBMV = tobacco vein banding mosaic virus; HC-Pro = helper component-proteinase; P3 = third protein; CP = coat protein; NIa = nuclear inclusion body 'a' protein; ORF = open reading frame

The objective of this study was to obtain more data about the subgrouping using more sequences. Ten new complete nucleotide and deduced amino acid sequences of TVBMV were collected from major tobacco production regions in China; these were compared with published sequences of TVBMV to determine the evolutionary relationship as well as genetic diversity of TVBMV isolates and to identify the most appropriate genome sequence with which to generate tobacco cultivars.

Materials and Methods

Virus isolates. Tobacco leaves with vein banding and yellow mosaic symptoms were extracted from Yishui (YS), Yinan (YN), Juxian (JX), Feixian (FX), Junan (JN), Zhucheng (ZC), Pingyi (PY), Changle (CL), Laiwu (LW), and Yiyuan (YY) regions in Shandong, China. Ten TVBMV isolates were screened using serological characterization and by amplifying a TVBMV CP conserved region of 200 bp. The CP1 (5'-GATGCACAAATGGAGATTGTTTTGA-3') and CP2 (5'-GAGTGACAACAGCCTCAGCGGTTGTTG-3') primers were used. The isolates were named YS, YN, JX, FX, JN, ZC, PY, CL, LW, and YY, corresponding to their origin. After purification by three inoculations through *Chenopodium amaranticolor*, isolates were mechanically inoculated into tobacco plants (*Nicotiana tabacum* cv. Samsun) in a greenhouse at approximately 25°C.

RT-PCR and determination of 10 complete sequences. Total RNA was extracted from 100-mg leaf tissues using Trizol reagent and cDNA was obtained with a RevertAid First Strand cDNA Synthesis Kit following the manufacturer's instructions (MBI Fermentas, Germany). Five pairs of degenerate primers (Table 1) were designed according to previously reported full-length genome sequences of TVBMV-YND and TVBMV-HN39 (Wang *et al.*, 2010; Yu *et al.*, 2007) to construct the entire genome sequences of the 10 TVBMV isolates. The 5'-end of the RNA genome was amplified by 5'-RACE using a 5'-Full RACE Kit (Takara, China). The expected amplified products were purified from 1.5% agarose gel using a gel extraction kit (Takara, China) and cloned into the pMD18-T vector

(Takara, China). The reconstructed vectors were transformed into *Escherichia coli* DH5 α competent cells according to the standard transformation method. At least two independent clones of each fragment were sequenced in both directions and the accurate nucleotide sequences were determined by comparisons.

Phylogenetic analysis. Sequences were assembled using DNAMAN (Lynnon Biosoft, Canada) and DNASTAR (DNASTAR Inc., USA). The complete genome sequences of YS, YN, JX, FX, JN, ZC, PY, CL, LW, and YY have been deposited to GenBank under Acc. Nos HQ396791, JN630468, JN630471, HQ396792, JN621319, HQ396793, JN630469, JN630470, JN630473, and JN630472. Nucleotide sequence alignments were performed using CLUSTAL W (Thompson *et al.*, 1994). Phylogenetic analysis of the complete genomes of the 10 isolates with YND and HN39 were conducted by neighbor joining, minimum evolution, and maximum parsimony in MEGA 4.1 software. Genetic distances within and between groups were estimated using the Pamilo-Bianchi-Li method (Pamilo and Bianchi, 1993).

Recombination analysis. The complete sequence alignments were subjected to recombination analysis with the RDP Software package (Martin *et al.*, 2010). Possible recombination events were detected using RDP, GENECONV, BOOTSCAN, MAXCHI, CHIMAERA, 3SEQ, and SISCAN packaged in RDP with the default settings and a Bonferroni-corrected *P* value cutoff of 0.05. Recombination events supported by at least four programs with *P* values lower than 1.0×10^{-6} were considered "clear"; otherwise, they were regarded as "tentative" ones.

Analysis of selection pressure and population demographics of coding genes. Selection pressure was evaluated using nonsynonymous (d_N) and synonymous (d_S) substitutions with Pamilo-Bianchi-Li (PBL) method (Pamilo and Bianchi, 1993; Li, 1993). Confidence estimates for d_N and d_S values were calculated using the bootstrap method with 500 replicates. A d_N/d_S ratio lower than 1 indicated that negative selection had occurred, a d_N/d_S ratio higher than 1 indicated positive selection and a d_N/d_S ratio of 1 indicated neutral selection. In addition, all the coding genes were subjected to sequence polymorphism using DnaSP 5.0 (Librado and Rozas, 2009) in calculating Tajima's *D*, Fu and Li's *D*, Fu and Li's *F*, the haplotype diversity and the nucleotide diversity to estimate their distributive conditions. Mismatch distribution was also estimated and the ragged, multimodal distribution in the samples suggested the long-term stability of the population; otherwise, the population would have evolved under expanding conditions.

Table 1. Primers used for TVBMV complete genomic sequence amplification

Primer name	Position (bp)	Nucleotide sequence (5' to 3')
TVBMV1-5	1–21	ATACGAAAAACGAACAAAGCA
TVBMV1-3	2479–2509	ACTGTATGAGCAACTTGACA
TVBMV2-5	2349–2368	ACAGGATACCACATCTTGAA
TVBMV2-3	3689–3670	TGTTTCATTCGACTCATCGA
TVBMV3-5	3609–3629	ATAGTTGGTGTTCAGATCAA
TVBMV3-3	5789–5770	TGCATCACGTGCATTCCTGA
TVBMV4-5	5671–5690	CTGGAGGTGCATGGATGATA
TVBMV4-3	7091–7072	GCTTGCACTACTGCTTTAA
TVBMV5-5	6984–7004	ACAAGCAAACATCATAGAAGAT
TVBMV5-3	9570–9551	CCCTCACACCAAATGATATT

Results

Sequence identity analyses

Sequence analysis showed that the complete genome RNA sequence of each isolate was 9570 nt long, excluding the poly(A) tail, comprising the 146-nt 5'-UTR, the 184-nt 3'-UTR, and a large ORF (nucleotides 147–9386) encoding

one large polyprotein. The 10 isolates had similar base compositions of the entire genome: adenine, 31.66%–31.84%; uracil, 26.33%–26.65%; cytosine, 18.84%–19.18%, and guanine, 22.70%–22.83%. Consistent with previously reported data for YND and HN39, the 5'-UTR of the 10 isolates contained one or two highly conserved potybox b (UCAAGCA) motifs but no potybox a (AUAACAU). For putative polyproteins, the cleavage sites of P1, HC-Pro, and NIa-Pro were Y/S, G/G, Q/A, Q/S, Q/S, Q/G, E/A, Q/S, and Q/G, identical with those of YND and HN39, except that YND had a unique NIb/CP cleavage site (Q/N). Unlike many potyviruses, the 12 TVBMV isolates had an RITC motif in HC-Pro, instead of a KITC motif, involved in aphid transmission.

Comparisons of nucleotide and amino acid sequences revealed that the YS-YY isolates shared nucleotide and amino acid identities of 96.00%–99.47% and 98.21%–99.61%, whereas with YND and HN39 shared those of 89.82%–97.45% and 95.03%–99.19%. The sequence identities of every region in the whole genome available in the GenBank were further analyzed with those described herein. For each region, all the isolates shared much lower identities with Yunnan isolates. The 3'-UTR was more conservative with the highest nucleotide identity (94.05%–100%), whereas the 5'-UTR had the lowest nucleotide identity (83.56%–100%). As for amino acid identities, the P1 protein had the most variable and least conserved region (82.14%–100%) in the entire genome.

Phylogenetic analyses

The complete genome sequences of the 10 TVBMV isolates were subjected to phylogenetic analysis with YND and HN39 (Fig. 1). The 12 TVBMV isolates were clustered into two groups, with YND forming a separate branch. The phylogenetic trees constructed with every region of their whole genomes yielded the same result, indicating that TVBMV sequence correlated with geographical origin.

Phylogenetic trees of the HC-Pro, P3, 6K1, and CP genes were constructed using all the available sequences to strengthen the evidence further. The phylogenetic analysis of HC-Pro, P3, 6K1, and CP (Fig. 2a–d) demonstrated that TVBMV was divided into geographical origin-specific subgroups. The nucleotide sequences of HC-Pro, P3, and 6K1 were grouped into two main branches: group I comprised all the isolates except the Yunnan isolates and group II consisted

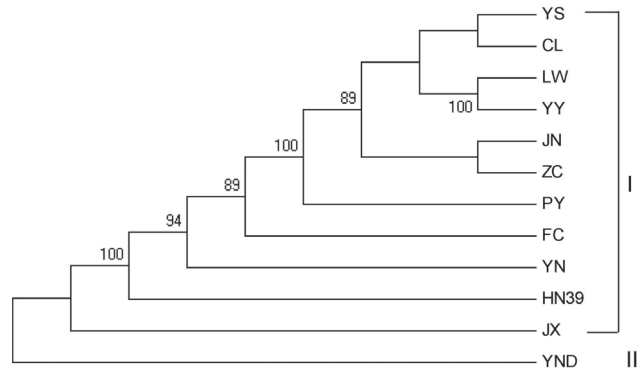


Fig. 1

Phylogenetic tree of TVBMV isolates constructed with the complete nucleotide sequence

Bootstrap analysis with 1000 replicates was performed. Branches with bootstrap values lower than 50% were collapsed and only bootstrap values >70% were shown.

of the latter set. The phylogenetic tree of the CP gene was divided into three groups: most isolates formed group I; the isolates from Japan, United States, and Taiwan constituted group II; and the Yunnan isolates comprised group III. Group I could be divided into three subgroups for HC-Pro, whereas it was divided into two subgroups for P3, 6K1, and CP. The within-group and between-group genetic distances were calculated to confirm the rationality of the subgrouping. The results showed that for the four genes, the genetic distances between groups were all one to four times higher than those within groups. All the statistical data showed that the findings from the phylogenetic analysis were reasonable and that geographical distribution is a potentially important selection factor.

Recombination analyses

The 10 isolates were included in RDP to identify possible recombination events. The results showed that several isolates were predicted to have recombination events, but only the sites found in YN, ZC, and PY were obvious (Table 2). The recombination sites were at the 3'-end of CI, the 5'-end of NIb, the 3'-end of HC-Pro, the 5'-end of VPg and the central region of 6K1. Recombination events between different groups were not found.

Table 2. Recombination message detected by RDP3

Isolates	Supporting software*	Major parent	Minor parent	P-value	Z-value	Recombination site
YN	RGBMS3	YY	YS	7.448×10^{-14}	6.39	5018–7101 nt
PY	RGBMS3	ZC	YN	5.645×10^{-27}	6.23	2353–5774 nt
ZC	RGBMCS3	JN	CL	1.555×10^{-11}	6.23	2431–3573 nt

*R: RDP; G: GENECONV; B: Bootscan; M: MaxChi; C: Chimaera; S: SiScan; 3: 3Seq. The software showed the highest significant P-value.

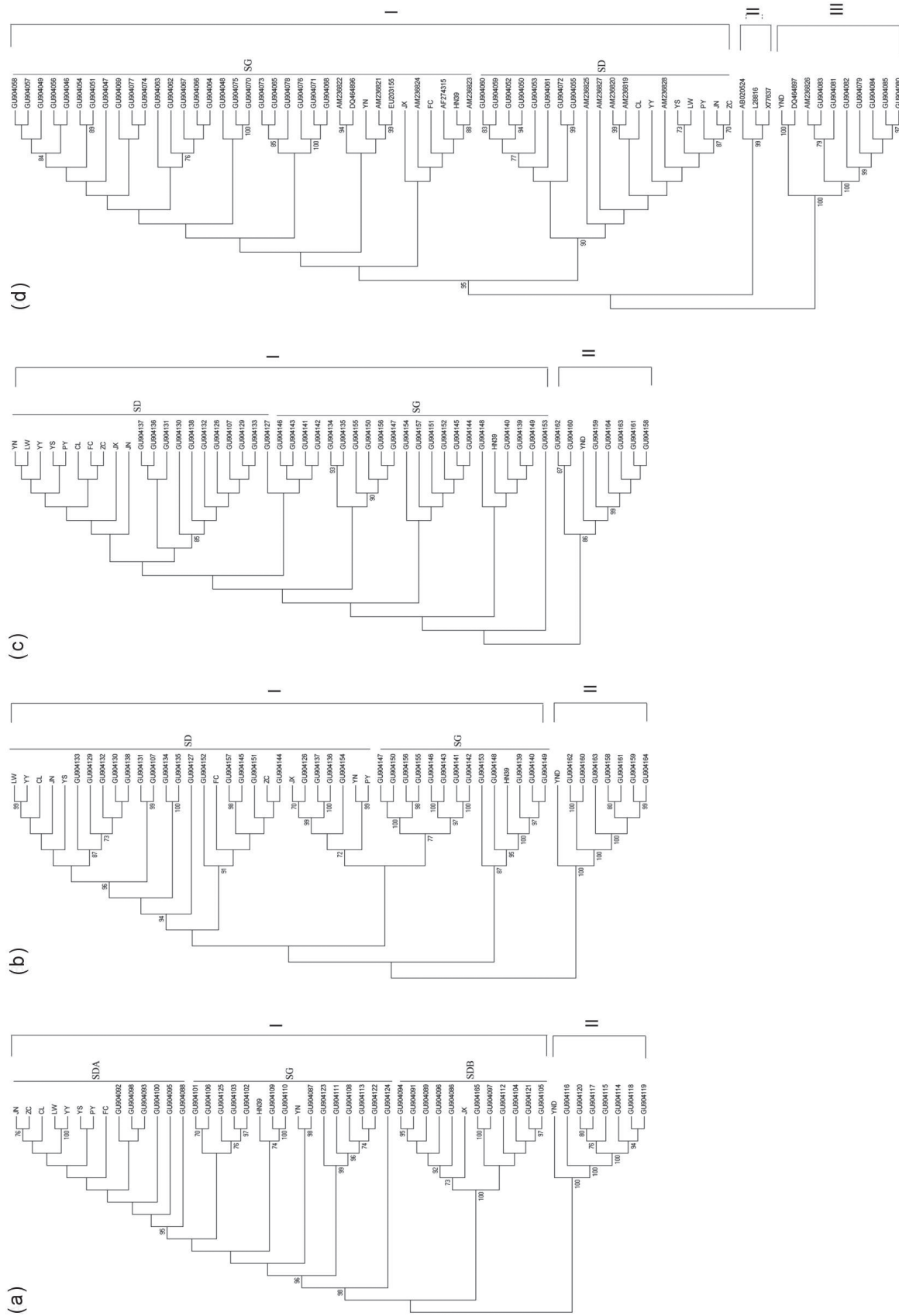


Fig. 2
Phylogenetic analysis of TVBMV isolates constructed with the HC-Pro, P3, 6K1, and CP nucleotide sequences

Bootstrap analysis with 1000 replicates was performed. Branches with bootstrap values lower than 50% were collapsed and only bootstrap values >70% were shown.

Genetic diversity and population demography

Coding genes were subjected to genetic diversity analysis to estimate their variability. Although, HC-Pro and NIa showed the highest (0.0545) and lowest (0.0303) values respectively, no clear difference between the genes was detected. The d_N/d_S ratio for the ORF was also calculated with the values for all the coding genes being lower than 1.0, suggesting that the ORF of TVBMV was under negative selection. On the other hand, the d_N/d_S ratio for P1 was considerably high, showing that the P1 gene was under the highest selection pressure. The values obtained for Tajima's D, Fu and Li's D, as well as Fu and Li's F of each coding gene were negative, but the *P* values were higher than 0.05 or 0.10, rendering the results inconclusive. The frequency distributions of the number of pairwise nucleotide differences obtained from the P1-CP sequences were all ragged and multimodal, indicating the long-term stability of the population.

Discussion

This study determined the complete genomes of 10 TVBMV isolates from China. Nucleotide and amino acid comparisons showed that the Yunnan isolates shared much lower identities with other isolates. Phylogenetic analysis also suggests that the Yunnan isolates constituted a specific group. Zhang *et al.* (2011) reported phylogenetic results showing that TVBMV is divided into three groups for HC-Pro (SD, MC, and YN) and CP, into two groups for P3 and 6K1, whereas we obtained novel results with the subgroups for HC-Pro, P3, and 6K1 by including more sequences in the current study. Based on HC-Pro, TVBMV was divided into two major groups. Group I, which was divided into three subgroups, corresponded to the formerly recognized SD and MC. For P3 and 6K1, group I was clustered into two subgroups; the previously identified group I could not be further divided. Overall, the phylogenetic trees demonstrated that the clustering of TVBMV correlated with geographical origin and that the isolates from mainland China, excluding the Yunnan ones, formed a large group.

Previous research has shown that the "hot" recombination spots of TVBMV occurred in the 3'-end of HC-Pro and the central region of 6K1. Recombination events in the 5'-end of HC-Pro, the 5'-end of P3 and in the CP gene were found also. In the current study we identified new recombination sites in the 3'-termini of HC-Pro and CI as well as the 5'-termini of VPg and NIb. Recombination was not detected between TVBMV isolates of different geographical groups.

For the TVBMV population, the d_N/d_S ratios for each coding gene were all lower than 1.0, indicating that the ORFs were under negative selection to preserve their protein functions. As the degree of selection pressure on different genes

was associated with the functions of their encoding proteins, different genes endured different selection pressures. The HC-Pro gene was under the strongest pressure, while on the other hand, the NIa gene was under the weakest pressure.

Mismatch distribution results implied that TVBMV had existed stably over an extended period of time. As detected in this study, there are two possible reasons behind its increasing incidence in recent years, firstly, TVBMV has always induced similar systems with PVY in fields, making PVY responsible for the great losses caused by TVBMV, and secondly, favorable environments enable TVBMV to spread quickly.

The lack of TVBMV-resistant tobacco (Zhang *et al.*, 2011) evidently prompts its immediate cultivation. Advances in the genetic transformation of crop plants and deeper understanding of the genomic components of TVBMV have allowed for the development of transgenic tobacco. Viral coat protein, replicase and movement protein genes have been typically used to produce virus-resistant crops (Dasgupta *et al.*, 2003; Batuman *et al.*, 2006); however, the 3'-UTR of viruses, which may obtain the same or an even more durable and stable level of resistance (Vaslin *et al.*, 2001; Batuman *et al.*, 2006), has been widely unexplored because its secondary structure is greatly associated with virus replication (Li *et al.*, 2006). As the identity of the 3'-UTR is higher and more stable than that of other regions, it may be used to produce TVBMV-resistant tobacco.

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References

- Batuman O, Mawassi M, Bar-Joseph M (2006): Transgenes consisting of a dsRNA of an RNAi suppressor plus the 3' UTR provide resistance to Citrus tristeza virus sequences in *Nicotiana benthamiana* but not in citrus. *Virus Genes* 33, 319–327.
- Chang BY, Huang CR, Yeh SD, Chiang JK, Hung LM, Hu HY (1994): Nucleotide-sequence of the coat protein coding region of the potyvirus tobacco vein-banding mosaic virus. *Arch. Virol.* 138, 17–25. <http://dx.doi.org/10.1007/BF01310035>
- Chin WT (1966): A survey of tobacco mosaic viruses in central Taiwan. *J. Agric. Ass. Chin.* 55, 85–88.
- Chin WT (1975): Symptomatological study of Tobacco vein banding mosaic virus on tobacco plant. *Bull. Taiwan Tob. Res. Inst.* 3, 65–70.
- Dasgupta I, Malathi VG, Mukherjee SK (2003): Genetic engineering for virus resistance. *Curr. Sci.* 84, 341–354.
- Habera LF, Berger PH, Reddick BB (1994): Molecular evidence from 3'-terminus sequence-analysis that Tobacco vein-banding mosaic virus is a distinct member of the Potyvirus group. *Arch. Virol.* 138, 27–38. <http://dx.doi.org/10.1007/BF01310036>

- Librado P, Rozas J (2009): DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25, 1451–1452. <http://dx.doi.org/10.1093/bioinformatics/btp187>
- Li XD, Yu XQ, Guo QS, Liu JL, Zhu XP, Guo XQ (2006): Advances on functional genomics of potyviruses. *Shandong Sci.* 19, 1–6.
- Li WH (1993): Unbiased estimation of the rates of synonymous and non-synonymous substitution. *J. Mol. Evol.* 36, 96–99. <http://dx.doi.org/10.1007/BF02407308>
- Martin DP, Lemey P, Lott M, Moulton V, Posada D, Lefevre P (2010): RDP3: a flexible and fast computer program for analyzing recombination. *Bioinformatics* 26, 2462–2463. <http://dx.doi.org/10.1093/bioinformatics/btq467>
- Moreno IM, Malpica JM, Díaz-Pendo'n JA, Moriones E, Fraile A, García-Arenal F (2004): Variability and genetic structure of the population of watermelon mosaic virus infecting melon in Spain. *Virology* 318, 451–460. <http://dx.doi.org/10.1016/j.virol.2003.10.002>
- Pamilo P, Bianchi NO (1993): Evolution of the Zfx and Zfy genes: Rates and interdependence between the genes. *Mol. Biol. Evol.* 10, 271–281.
- Reddick BB, Collins-Shepard MH, Christie RG, Gooding GV (1992): A new virus-disease in North-America caused by Tobacco vein-banding mosaic virus. *Plant Dis.* 76, 856–859. <http://dx.doi.org/10.1094/PD-76-0856>
- Thompson JD, Higgins DG, Gibson TJ (1994): CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positions-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 22, 4673–4680. <http://dx.doi.org/10.1093/nar/22.22.4673>
- Tian YP, Liu JL, Yu XQ, Lei LP, Zhu XP, Valkonen JPT, Li XD (2007): Molecular diversity of Tobacco vein banding mosaic virus. *Arch. Virol.* 152, 1911–1915. <http://dx.doi.org/10.1007/s00705-007-1020-3>
- Tomitaka Y, Ohshima K (2006): A phylogeographical study of the Turnip mosaic virus population in East Asia reveals an emergent lineage in Japan. *Mol. Ecol.* 15, 4437–4457. <http://dx.doi.org/10.1111/j.1365-294X.2006.03094.x>
- Tsompana M, Abad J, Purganan M, Moyer W (2005): The molecular population genetics of the Tomato spotted wilt virus (TSWV) genome. *Mol. Ecol.* 14, 53–66. <http://dx.doi.org/10.1111/j.1365-294X.2004.02392.x>
- Vaslin MF, Vidal MS, Alves ED, Farinelli L, de Oliveira DE (2001): Co-suppression mediated virus resistance in transgenic tobacco plants harboring the 3'-untranslated region of Andean potato mottle virus. *Transgenic Res.* 10, 489–499. <http://dx.doi.org/10.1023/A:1013015401546>
- Wang HY, Zhu TS, Cui TT, Hou SS, Yin X, Li XD, Lei LP, Zhu XP (2010): Complete genome sequence of a tobacco isolate of the Tobacco vein banding mosaic virus strain prevailing in China. *Arch. Virol.* 155, 293–295. <http://dx.doi.org/10.1007/s00705-009-0570-y>
- Yu XQ, Lan YF, Wang HY, Liu JL, Zhu XP, Valkonen JPT, Li XD (2007): The complete genomic sequence of Tobacco vein banding mosaic virus and its similarities with other potyviruses. *Virus Genes* 35, 801–806. <http://dx.doi.org/10.1007/s11262-007-0135-7>
- Zhang CL, Gao R, Wang J, Zhang GM, Li XD, Liu HT (2011): Molecular variability of Tobacco vein banding mosaic virus populations. *Virus Res.* 158, 188–198. <http://dx.doi.org/10.1016/j.virusres.2011.03.031>
- Zhang ZK, Li Y (2001): *Plant Viruses in Yunnan Province*. Scientific Press, Beijing, pp. 46–49.