

Lymphatic vessel density and expressions of lymphangiogenic growth factors in salivary carcinomas

M. F. MELLO¹, A. F. COSTA¹, L. L. FREITAS¹, A. B. SOARES², V. C. ARAUJO², A. J. TINCANI³, A. S. MARTINS³, A. ALTEMANI¹

¹Department of Pathology, School of Medicine, State University of Campinas (UNICAMP), Campinas, São Paulo, Brazil, e-mail: aaltemani@uol.com.br, ²Department of Oral Pathology, São Leopoldo Mandic Institute and Research Center, Campinas, São Paulo, Brazil, ³Head and Neck Service, Department of Surgery, School of Medicine, State University of Campinas (UNICAMP), Campinas, São Paulo, Brazil.

Received October 28, 2010

Nodal metastasis is an important prognostic indicator in head and neck cancers, including salivary carcinomas. In these, the risk for lymph node metastasis is variable and strongly associated with the tumor histologic type. The aim of the current study was to evaluate the lymphatic vessel density (LVD) and expressions of lymphangiogenic growth factors by tumor cells in different histologic types of salivary carcinomas subdivided according to the risk for nodal metastasis. In 15 high-risk (undifferentiated, high-grade mucoepidermoid and salivary duct carcinomas) and 60 low/moderate-risk tumors (adenoid cystic, low/intermediate-grade mucoepidermoid, acinic cell, myoepithelial, epithelial-myoepithelial and polymorphic low-grade carcinomas) the expressions of vascular endothelial growth factor-C (VEGF-C), hepatocyte growth factor (HGF) and D2-40 (for assessing LVD) were examined. No significant differences were encountered between high- and low/moderate/-risk carcinomas regarding LVD and VEGF-C or HGF expressions. Furthermore, the expression of these proteins did not correlate with LVD. Lymphatic vascular invasion was found mainly in high-risk carcinomas. Intratumoral LVD was significantly lower than peritumoral, regardless of the risk for metastasis and primary site of the lesion. The histologic types of salivary carcinomas which are associated with high-risk for nodal metastasis do not present increased LVD or VEGF-C and HGF expressions. The greater tendency for metastasis in these carcinomas seems to be related to their capacity to invade lymph vessels. Further studies on tumor cell interactions with lymphatic endothelial cells are needed to improve our understanding of the metastatic potential of salivary carcinomas.

Key words: lymph vessels, salivary carcinoma, VEGF-C, HGF.

Nodal metastasis is an important prognostic predictor in head and neck cancers, including salivary carcinomas. The risk for lymph node metastasis in these carcinomas is highly variable (9% – 85%) [1] and strongly associated with the tumor histologic type [1-5]. It is a general consensus that high-grade mucoepidermoid carcinomas, salivary duct carcinomas, undifferentiated carcinomas and squamous cell carcinomas are high-risk tumors for nodal metastasis [1, 3-5]. In these the risk for neck metastasis has been described to be >50% [1] and, thus, elective neck dissection is considered in their management [1-3].

Lymph vessels provide the main avenue for nodal metastasis and in head and neck squamous cell carcinoma (HNSCC) it has been shown that tumors of different anatomic regions do not vary significantly in their lymphangiogenic properties [6, 7]. However, a high lymph vessel density (LVD) seems to be an indicator of the risk of lymph node metastasis in HNSCC

[6-9]. Regarding salivary gland tumors, the lymphatic vessels have rarely been studied [10-12] and in a particular type of salivary carcinoma, i.e. in those arising in pleomorphic adenomas (CXPA) the lymphatic network was found to be composed mainly of pre-existing vessels [10].

The aim of the current study was to evaluate LVD in different histologic types of salivary carcinomas subdivided according to the risk for nodal metastasis. Furthermore we looked at the clinical follow up of the patients and analyzed expressions of lymphangiogenic growth factors by tumor cells. Lymphangiogenesis is regulated by multiple growth factors [13] in which vascular endothelial growth factor (VEGF)-C plays a key role as an essential and selective lymphangiogenic factor whereas hepatocyte growth factor (HGF) is a novel lymphangiogenic factor with an indirect mechanism of action [14]. The contribution of these proteins to tumor LVD and nodal metastasis in salivary carcinomas has yet to be investigated.

Materials and methods

The present study was approved by the Committee of Ethics of the University of Campinas, Brazil and was performed in 75 cases of salivary carcinomas which were retrieved from the files of the Department of Pathology of the University of Campinas. The study population consisted of 30 men (40%) and 45 women (60%), the average age at the time of diagnosis was 52.71 years (ranged from 7 to 72 years). The tumors were classified according to Regis de Brito Santos et al (2001) in low / moderate-risk for nodal metastasis [1] – 60 cases (adenoid cystic carcinomas – 15 cases, low / intermediate-grade mucoepidermoid carcinomas – 8, acinic cell carcinomas – 13, myoepithelial carcinomas- 6, epithelial-myoepithelial carcinomas- 12 and polymorphic low-grade adenocarcinomas- 6) and high-risk – 15 cases (undifferentiated carcinomas – 3 cases, high-grade mucoepidermoid carcinomas – 5 and salivary duct carcinomas 7). Regarding tumor location 64.71% were in the major salivary glands and 35.29% in the minor ones. Demographic and clinical information was obtained from the patients' medical records. None of them had received preoperative chemotherapy or radiotherapy.

Tissue microarrays. Salivary carcinoma samples were selected for tissue microarray (TMA) studies to evaluate the expressions of VEGF-C and HGF in tumor cells by immunohistochemical method. Three tissue cores of 1mm diameter were punched (Tissue-Tek-Quick-Ray™ Tissue Microarray System; Sakura; USA) as representative of the whole tumor in each case. Necrotic or hemorrhagic areas were avoided. The selected cores were placed into receptor blocks of TMA. Sections with 5µm thickness were obtained for the immunohistochemical studies.

Immunohistochemistry The antibodies used in this study were D2-40 (for detection of lymph vessels), VEGF-C and HGF (Table 1). For immunohistochemical staining, 5µm sections from each paraffin block were deparaffinized, hydrated and endogenous peroxidase activity was quenched by immersion of the slides in 3% hydrogen peroxide. The antigen retrieval (AR) was achieved by boiling them in a steamer immersed in citrate buffer (pH 6.0), except for D2-40. For D2-40 AR was performed using TrisEDTA (pH 9.0). After washing, the sections were incubated at 4°C, with the primary antibody, overnight. Signal detection was performed using EnVision peroxidase system (DAKO, Carpinteria, CA, USA) for 1h at 37°C. Subsequently, sections were stained for 5 min at 37°C with 3.3'-diaminobenzidine tetrahydrochloride (DAB) and counter-stained with Mayer's

hematoxylin. The isotype-matched negative controls did not show colored-precipitate on the tissue, which indicates that artifactual staining was minimized.

Evaluation of staining. Assessment of LVD: Intratumoral LVD (3 hotspots were located within tumor mass) and peritumoral LVD (3 hotspots were located within an area of 500µm from the tumor border) were assessed separately. In each tumor section stained for D2-40 the lymphatic vessels were manually counted at 200X (0.7386 mm² field) by two authors (MFM and LF) using a double-headed microscope. The mean number of lymphatics assessed was determined as LVD and the mean peritumoral LVD and intratumoral LVD were calculated for each case. Invasion of the D2-40 intratumoral or peritumoral lymphatic vessels by carcinoma cells (tumor emboli) was also evaluated.

Assessment of VEGF-C and HGF expressions: Brown staining of the cell cytoplasm was regarded as positive. The relative numbers of neoplastic VEGF-C+ and HGF+ cells were considered in relation to all neoplastic cells observed in each stained section. When the number of positive cells was more than 10%, the case was judged positive. The positive neoplastic cells were assessed regarding quantity using a three-tiered scale: >10% – 25%, >25% – 50% and > 50% of positive cells.

Statistical analysis. A chi-square test or a Fisher exact test was used to assess the associations among categorical data. Kruskal-Wallis test and Mann-Whitney U test were used for comparison of the numeric variables between the different groups as appropriate. Data were presented as mean + SD, and the results with $p < 0.05$ were considered significant. All the statistical procedures were performed using SPSS software for Windows, version 12.0 (SPSS® Inc. Illinois, USA).

Results

LVD. Peritumoral LVD was significantly higher than intratumoral LVD (Fig 1) in carcinomas with high-risk for nodal metastasis as well as in low/moderate-risk tumors. In the latter, peritumoral LVD means ranged from 15.83 to 71.83 and intratumoral from 0.50 to 12.40. No significant difference was found among the carcinomas of this group ($p = 0.130$ for peritumoral LVD and $p = 0.209$ for intratumoral). In high-risk tumors peritumoral LVD means ranged from 13.33 to 39.71 and intratumoral from 0.00 to 8.40 and no significant difference was found among these carcinomas either ($p = 0.311$ for peritumoral LVD and $p = 0.233$ for intratumoral). Furthermore, no correlation was detected between intratumoral and peritumoral LVD and risk for nodal metastasis or tumor site (Table 2).

Tumor emboli within lymph vessels were observed in 1 out of 60 cases (1.6%) in low/moderate-risk carcinomas and in 3 out of 15 (20%) in high-risk tumors. These emboli (Fig 1F) were found in intratumoral (4 cases) and peritumoral (1 case) lymphatics (one high-grade mucoepidermoid carcinoma presented intra and peritumoral emboli). Nodal metastasis at time of primary surgery was detected in 5 patients and their

Table 1. Details of the antibodies used for immunohistochemistry.

Antibody	Clone	Dilution	Source	Buffer (antigen retrieval)
D2-40	D240	1:200	*DAKO	Tris-EDTA
VEGF-C	H-190	1:100	# Santa Cruz	Citrate
HGF	N-19	1:500	# Santa Cruz	Citrate

* Dako Corporation Glostrup, Denmark; # Santa Cruz Biotechnology, USA.

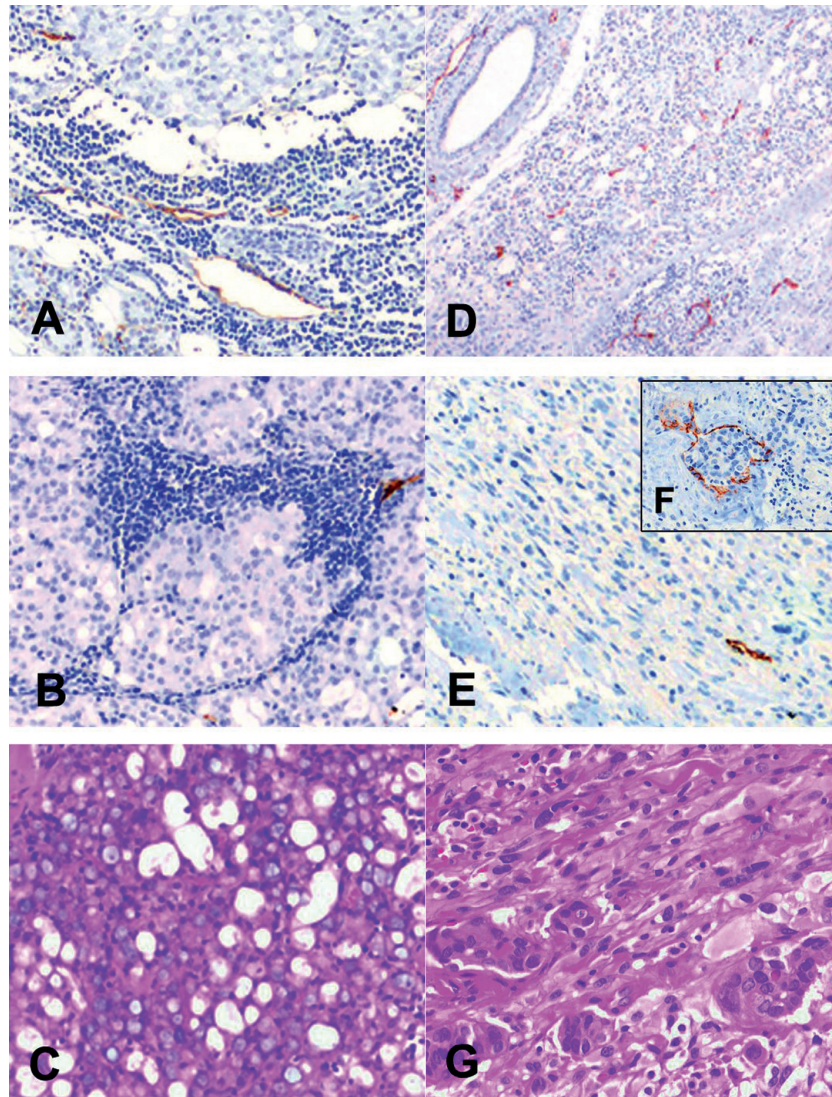


Figure 1 – Peritumoral and intratumoral lymphatic vessels stained for D2-40 (brown): A, B and C - acinic cell carcinoma (low/ moderate-risk tumor for nodal metastasis); D, E, F and G - salivary duct carcinoma with sarcomatoid component (high-risk tumor for nodal metastasis). The peritumoral lymphatic vessel density (A, D) is higher than the intratumoral one (B, E) in both types of carcinomas. Note the tumor embolus within an intratumoral lymphatic vessel in the salivary duct carcinoma (inset - F).

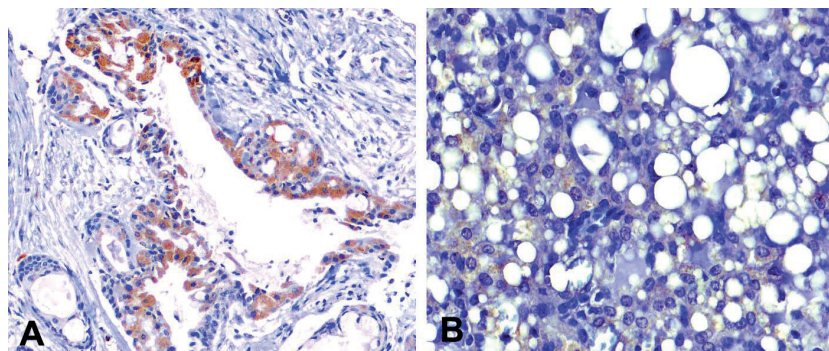


Figure 2 – VEGF-C (A) and HGF (B) expressions by tumor cells in mucoepidermoid and acinic cell carcinomas.

Table 2. Correlations of LVD with clinicopathological parameters and VEGF-C and HGF expressions.

Factors	N	ILVD (mean + SD)	P	PLVD (mean + SD)	P	*p
Risk for metastasis			0.953		0.518	
Low/moderate	60	7.90+17.43		43.54+45.08		0.000
High	15	5.17+8.03		33.67+29.31		
Tumor location			0.766		0.927	
Major salivary glands	33	9.64+18.92		39.53+32.48		0.000
Minor salivary glands	24	7.38+16.76		49.46+60.14		
Nodal Metastasis			0.608		0.806	
With	5	9.40+18.82		46.00+51.73		0.000
Without	55	6.15+14.51		38.50+43.52		
VEGF-C expression by tumor cells			0.774		0.319	
≤ 25%	29	6.93+16.19		39.34+28.38		0.000
> 25%	16	4.50+10.75		34.75+38.12		
HGF expression by tumor cells			0.275		0.328	
≤ 25%	30	7.43+16.39		38.00+29.08		0.000
> 25%	13	4.00+11.82		34.69+39.82		

*Intratumoral versus peritumoral lymphatic vessel density. ILVD, intratumoral lymphatic vessel density; PLVD, peritumoral lymphatic vessel density.

Table 3. Expressions of VEGF-C and HGF in salivary carcinomas subdivided according to the risk for nodal metastasis.

Growth factor	Total cases	Low/moderate-risk	High-risk	p
VEGF-C expression by tumor cells				0.612
≤ 25%	13	20.0%	18.8%	
> 25%	53	80.0%	81.2%	
HGF expression by tumor cells				0.191
≤ 25%	46	62.7%	81.8%	
> 25%	24	37.3%	18.2%	

tumors did not show significant differences in terms of LVD when compared with those of patients without metastasis (Table 2).

VEGF-C and HGF expressions. No correlation was found between the amount of positive cells for VEGF-C and HGF (Fig 2) and intratumoral or peritumoral LVD (Table 2). The expression of these proteins did not correlate with the risk for nodal metastasis either (Table 3).

Discussion

To our knowledge, this is the first study in which LVD was assessed in a series of different histologic types of salivary carcinomas from diverse primary locations (major and minor salivary glands). We showed that high-risk tumors for nodal metastasis (high-grade mucoepidermoid carcinomas, salivary duct carcinomas, undifferentiated carcinomas) presented no significant difference in LVD when compared with low/moderate-risk carcinomas. These findings suggest that high- and

low-risk salivary carcinomas are equally lymphangiogenic. Furthermore, despite the small number of patients with nodal metastasis, the absence of significant differences of LVD between salivary carcinomas with and without metastasis reinforces this assumption. Therefore, measurements of LVD seem to have no utility for estimation of metastatic risk for salivary carcinomas.

Our findings contrast with the observations for HNSCCs, particularly for SCC affecting the oral cavity, which is also a common site for salivary carcinomas [7, 8, 15]. In HNSCC, LVD measurements have been suggested as potential indicators of the risk of lymph node metastasis [6-9]. We believe that these discrepancies between SCC and salivary carcinomas regarding LVD and risk for nodal metastasis may reflect genuine differences between the malignant behaviors of these two types of carcinomas of the head and neck region. The mechanisms by which tumor cells leave the primary site, invade lymphatics and metastasize are complex. We speculate that in salivary carcinomas the main difference between high- and low-risk tumors for nodal metastasis could be their capacity to invade lymph vessels in addition to the mere presence of these channels. In this sense, in the current series, the finding of tumor emboli mainly in high-risk carcinomas lends support to this hypothesis. The mechanisms that control tumor cell interactions with lymphatic endothelium are still poorly understood and further studies are needed to explore the genetic and biochemical determinants involved in tumor cell entry into lymphatics in salivary carcinomas.

In human cancers, LVD has been analyzed within the main neoplastic mass (intratumoral lymphatics) as well as at the tumor margin (peritumoral lymphatics). In certain types of neoplasms, such as breast, ovarian, endometrial and lung cancers [16-19], no intratumoral lymphatic network has been found. In contrast, in others such as cutaneous melanoma,

HNSCC and SCC of uterine cervix [8, 20, 21] intratumoral lymphatics do exist and have been associated with nodal metastasis and adverse clinical outcome. In salivary carcinomas we detected lymph vessels in both intra- and peritumoral regions, although the latter was significantly higher regardless of the tumor site and metastatic risk. These findings suggest that the density of lymph vessels is governed by factors unrelated to histologic differentiation or primary tumor location. It has been postulated that growing tumor cells mechanically collapse or destroy the lymphatic network [22] and this could explain the significantly lower intratumoral LVD in salivary carcinomas. Low intratumoral LVD has also been found in HNSCC [7, 23]. It is of interest is that in our cases tumor emboli were detected mainly in intratumoral lymphatics, strengthening our previous suggestion that these can act as conduits for tumor cells in salivary carcinomas [10].

VEGF-C and VEGF-D are ligands for the receptor tyrosine kinase VEGFR-3 that is expressed on lymphatic endothelium. In experimental models, it has been shown that overexpression of VEGF-C or -D induces lymph vessel growth and lymph node metastases [24-27]. However, in human tumors this correlation has not always been found. There are contradictory results in different cancers or even in the same type of tumor with regard to VEGF-C expression and lymphangiogenesis or nodal metastasis [6, 20, 28]. In the current series, carcinomas with high- and low/moderate-risk for nodal metastasis presented similar expressions of VEGF-C. Furthermore, no correlation between VEGF-C expression by carcinoma cells and number of intra/ peritumoral lymphatics was encountered. These findings suggest that in salivary carcinomas tumor-produced VEGF-C may not be functional or alternatively this protein is not the only vascular stimulator involved in tumor lymphangiogenesis. This process is regulated by multiple factors that are produced by various cell types, including tumor-associated macrophages and fibroblasts [20, 29, 30]. Moreover, in addition to VEGF-C/VEGF-D/VEGFR-3 axis (the main signal transduction system in lymphatics), other lymphangiogenic factors (such as HGF and angiopoietin) have recently been described to influence this system (reviewed in Van der Auwera et al 2006).

HGF is a growth factor that belongs to the plasminogen-prothrombin gene superfamily that has a direct role in promoting tumor cell growth and invasion and stimulates lymphangiogenesis through an indirect mechanism [14]. In tumor model overexpression of HGF was found to induce lymphatic vessel growth in the peritumoral region [14] and in human oral SCC a significant correlation was detected between LVD and HGF expression [31]. In contrast, in high grade salivary carcinomas absence of correlation between HGF expression in tumor cells and regional lymph node and distant metastasis has been reported [32]. Our findings are in agreement with these data as we did not observe any significant relationship between HGF tumor expression and LVD. However, stromal expression of HGF, which was not studied in this work, has been described to be related to metastasis in salivary carcinoma

[32]. Therefore, the role of HGF in metastasis of salivary carcinomas may involve mechanisms other than lymphangiogenesis that need to be better explored.

In conclusion, the histologic types of salivary carcinomas which are associated with high-risk for nodal metastasis do not present increased LVD or VEGF-C and HGF expressions. It is likely that the greater tendency for metastasis in these carcinomas depends on their capacity to invade lymph vessels.

Acknowledgements: This work was supported by Conselho Nacional de Desenvolvimento Científico e Tecnológico – CNPq – Brazil (grant number 473641/2008-2009) and Fundação de Amparo à Pesquisa do Estado de São Paulo – FAPESP – Brazil (Grant number 2007/55336-2).

References

- [1] RÉGIS DE BRITO SANTOS I, KOWALSKI LP, ARAUJO VC, LOGULLO AF, MAGRIN J. Multivariate analysis of risk factors for neck metastases in surgically treated parotid carcinomas. *Arch Otolaryngol Head Neck Surg* 2001; 127: 56–60.
- [2] KELLEY DJ, SPIRO RH. Management of the neck in parotid carcinoma. *Am J Surg* 1996; 172: 695–7. doi:10.1016/S0002-9610(96)00307-8
- [3] FERLITO A, SHAHA AR, RINALDO A, MONDIN V. Management of clinically negative cervical lymph nodes in patients with malignant neoplasms of the parotid gland. *ORL J Otorhinolaryngol Relat Spec* 2001; 63: 123–6. doi:10.1159/000055726
- [4] STENNERT E, KISNER D, JUNGEHUELING M, GUNTINAS-LICHIUS O, SCHRÖDER U et al. High incidence of lymph node metastasis in major salivary gland cancer *Arch Otolaryngol Head Neck Surg* 2003; 129: 720–3. doi:10.1001/archotol.129.7.720
- [5] LLOYD S, YU JB, WILSON LD, DECKER RH. A prognostic index for predicting lymph node metastasis in minor salivary gland cancer. *Int J Radiat Oncol Biol Phys* 2010; 76: 169–75. doi:10.1016/j.ijrobp.2009.01.021
- [6] BEASLEY NJ, PREVO R, BANERJI S, LEEK RD, MOORE J et al. Intratumoral lymphangiogenesis and lymph node metastasis in head and neck cancer. *Cancer Res* 2002; 62: 1315–20.
- [7] FRANCHI A, GALLO O, MASSI D, BARONI G, SANTUCCI M. Tumor lymphangiogenesis in head and neck squamous cell carcinoma. A morphometric study with clinical correlations. *Cancer* 2004; 101: 973–8. doi:10.1002/cncr.20454
- [8] KYZAS PA, GELEFF S, BATISTATOU A, AGNANTIS NJ, STEFANOUD. Evidence for lymphangiogenesis and its prognostic implications in head and neck squamous cell carcinoma. *J Pathol* 2005; 206: 170–7. doi:10.1002/path.1776
- [9] AUDET N, BEASLEY NJ, MACMILLAN C, JACKSON DG, GULLANE PJ, et al. Lymphatic vessel density, nodal metastases, and prognosis in patients with head and neck cancer. *Arch Otolaryngol Head Neck Surg* 2005; 131: 1065–70. doi:10.1001/archotol.131.12.1065
- [10] SOARES AB, PONCHIO L, JULIANO PB, ARAÚJO VC, ALTEMANI A. Lymphatic vascular density and lym-

- phangiogenesis during tumour progression of carcinoma ex pleomorphic adenoma. *J Clin Pathol* 2007; 60: 995–00. doi:10.1136/jcp.2006.042523
- [11] SOARES AB, ARAÚJO VC, JULIANO PB, ALTEMANI A. Angiogenic and lymphangiogenic microvessel density in recurrent pleomorphic adenoma. *J Oral Pathol Med.* 2009; 38: 623–9.
- [12] TEYMOORTASH A, SCHRADER C, SHIMODA H, KATO S, WERNER JA. Evidence of lymphangiogenesis in Warthin's tumor of the parotid gland. *Oral Oncol.* 2007; 43: 614–8. doi:10.1016/j.oraloncology.2006.07.006
- [13] CAO Y. Opinion: emerging mechanisms of tumour lymphangiogenesis and lymphatic metastasis. *Nat Rev Cancer* 2005; 5: 735–43. doi:10.1038/nrc1693
- [14] CAO R, BJÖRNDAHL MA, GALLEGRO MI, CHEN S, RELIGA P, HANSEN AJ, CAO Y. Hepatocyte growth factor is a lymphangiogenic factor with an indirect mechanism of action. *Blood* 2006; 107: 3531–6. doi:10.1182/blood-2005-06-2538
- [15] MIYAHARA M, TANUMA J, SUGIHARA K, SEMBA I. Tumor lymphangiogenesis correlates with lymph node metastasis and clinicopathologic parameters in oral squamous cell carcinoma. *Cancer* 2007; 110: 1287–94. doi:10.1002/cncr.22900
- [16] WILLIAMS CS, LEEK RD, ROBSON AM, BANERJI S, PREVO R et al. Absence of lymphangiogenesis and intratumoural lymph vessels in human metastatic breast cancer *J Pathol* 2003; 200: 195–06. doi:10.1002/path.1343
- [17] SCHOPPMANN SF, HORVAT R, BIRNER P. Lymphatic vessels and lymphangiogenesis in female cancer: mechanisms, clinical impact and possible implications for anti-lymphangiogenic therapies (Review). *Oncol Rep* 2002; 9: 455–60.
- [18] SCHMID K, BIRNER P, GRAVENHORST V, END A, GELEFF S. Prognostic value of lymphatic and blood vessel invasion in neuroendocrine tumors of the lung. *Am J Surg Pathol* 2005; 29: 324–8 doi:10.1097/01.pas.0000149706.74216.b6
- [19] KOUKOURAKIS MI, GIATROMANOLAKI A, SIVRIDES E, SIMOPOULOS C, GATTER KC et al. LYVE-1 immunohistochemical assessment of lymphangiogenesis in endometrial and lung cancer. *J Clin Pathol* 2005; 58: 202–6. doi:10.1136/jcp.2004.019174
- [20] DADRAS SS, PAUL T, BERTONCINI J, BROWN LF, MUZIKANSKY A, JACKSON DG et al. Tumor lymphangiogenesis: a novel prognostic indicator for cutaneous melanoma metastasis and survival. *Am J Pathol* 2003; 162: 1951–60. doi:10.1016/S0002-9440(10)64328-3
- [21] GOMBOS Z, XU X, CHU CS, ZHANG PJ, ACS G. Peritumoral lymphatic vessel density and vascular endothelial growth factor C expression in early-stage squamous cell carcinoma of the uterine cervix. *Clin Cancer Res* 2005; 11:8364–71. doi:10.1158/1078-0432.CCR-05-1238
- [22] PADERA TP, KADAMBI A, DI TOMASO E, CARREIRA CM, BROWN EB et al. Lymphatic metastasis in the absence of functional intratumoral lymphatics. *Science* 2002; 296: 1883–6. doi:10.1126/science.1071420
- [23] XUAN M, FANG YR, WATO M, HATA S, TANAKA A. Immunohistochemical co-localization of lymphatics and blood vessels in oral squamous cell carcinomas. *J Oral Pathol Med* 2005; 34: 334–9. doi:10.1111/j.1600-0714.2005.00316.x
- [24] KARPANEN T, EGEBLAD M, KARKKAINEN MJ, KUBO H, YLÄ-HERTTUALA S et al. Vascular endothelial growth factor C promotes tumor lymphangiogenesis and intralymphatic tumor growth. *Cancer Res* 2001; 61:1786–90.
- [25] MANDRIOTA SJ, JUSSILA L, JELTSH M, COMPAGNI A, BAETENS D et al. Vascular endothelial growth factor-C-mediated lymphangiogenesis promotes tumour metastasis. *EMBO J* 2001; 20: 672–82. doi:10.1093/emboj/20.4.672
- [26] SKOBE M, HAWIGHORST T, JACKSON DG, PREVO R, JANES L et al. Induction of tumor lymphangiogenesis by VEGF-C promotes breast cancer metastasis. *Nat Med* 2001; 7: 192–8. doi:10.1038/84643
- [27] STACKER SA, CAESAR C, BALDWIN ME, THORNTON GE, WILLIAMS RA et al. VEGF-D promotes the metastatic spread of tumor cells via the lymphatics. *Nat Med* 2001; 7: 186–91. doi:10.1038/84635
- [28] OHNO F, NAKANISHI H, ABE A, KINISHITA A, HASEGAWA Y et al. Regional difference in intratumoral lymphangiogenesis of oral squamous cell carcinomas evaluated by immunohistochemistry using D2-40 and podoplanin antibody: an analysis in comparison with angiogenesis. *J Oral Pathol Med* 2007; 36: 281–9. doi:10.1111/j.1600-0714.2007.00529.x
- [29] SCHOPPMANN SF, BIRNER P, STÖCKL J, KALT R, ULLRICH R et al. Tumor-associated macrophages express lymphatic endothelial growth factors and are related to peritumoral lymphangiogenesis. *Am J Pathol* 2002; 161: 947–56. doi:10.1016/S0002-9440(10)64255-1
- [30] VAN DER AUWERA I, CAO Y, TILLE JC, PEPPER MS, JACKSON DG et al. First international consensus on the methodology of lymphangiogenesis quantification in solid human tumours. *Br J Cancer* 2006; 95: 1611–25. doi:10.1038/sj.bjc.6603445
- [31] ALI MA. Lymphatic microvessel density and the expression of lymphangiogenic factors in oral squamous cell carcinoma. *Med Princ Pract* 2008; 17: 486–92. doi:10.1159/000151572
- [32] TSUKINOKI K, YASUDA M, MORI Y, ASANO S, NAITO H et al. Hepatocyte growth factor and c-Met immunoreactivity are associated with metastasis in high grade salivary gland carcinoma. *Oncol Rep* 2004; 12: 1017–21.